

ELISA ENZYME LINKED IMMUNOSORBENT ASSAY

Microwell Method

Rubella IgG



For in vitro Diagnostic Use

Product Insert

Enzyme Linked Immunosorbent Assay for the **qualitative** and **quantitative** determination of IgG Antibodies to Rubella Virus in human serum or plasma. It is intended as an aid in the diagnosis of possible Rubella infection.





Microwell Method - 96 wells
(12 x 8-well Antigen coated Strips
Individual breakaway)

INTRODUCTION

Rubella is a small spherical enveloped RNA virus belonging to Togaviridae family. Most commonly known as the German or 3-day measles, the Rubella virus is spread through droplet infection resulting in mild contagious rash in children or young adults. In childhood, the infection is self-limited, benign disease characterized by low-grade fever, headache, lymphadenopathy, arthralgia, and conjunctivitis. However, infection during pregnancy particularly in the first trimester can lead to spontaneous abortion, intrauterine infection causing fetal death, or congenital abnormalities. Congenital rubella depends on the time the infection occurs and may result in severe complications including deafness, ocular problems including cataracts and glaucoma, congenital heart disease and mental retardation.^{1,2} IgM antibodies against rubella are first produced reaching detectable levels within 2-3 days and peak 14-21 days after onset of symptoms which remain detectable over the next 4-8 weeks. Diagnosis of active or recent infection may be obtained by presence of IgM antibody in single early specimen. After several days, IgG antibodies appear after IgM response and peak 14-21 days later which then persist at varying levels for life.^{3,4} The presence of IgG antibodies to rubella is indicative of previous infection and presumptive immunity.^{5,6}

The Dialab Rubella IgG ELISA Test Kit is an immunoassay for the qualitative and quantitative detection of the presence of IgG antibodies to Rubella in serum or plasma specimen. The test utilizes purified Rubella antigens to selectively detect IgG antibodies to Rubella in serum or plasma.

PRINCIPLE OF THE ASSAY

The Dialab Rubella IgG ELISA Test Kit is a solid phase enzyme immunoassay based on indirect principle for the qualitative and quantitative detection of IgG antibodies to Rubella in human serum or plasma. The microwell plate is coated with Rubella antigens. During testing, the specimen diluent and the specimens are added to the antigen coated microwell plate and then incubated. If the specimens contain IgG antibodies to Rubella, it will bind to the antigens coated on the microwell plate to form immobilized antigen-Rubella IgG antibody complexes. If the specimens do not contain IgG antibodies to Rubella, the complexes will not be formed. After initial incubation, the microwell plate is washed to remove unbound materials. The enzyme-conjugated anti-human IgG antibodies are added to the microwell plate and then incubated. The enzyme-conjugated antihuman IgG antibodies will bind to the immobilized antigen-Rubella IaG antibody complexes present. After the second incubation, the microwell plate is washed to remove unbound materials. Substrate Solution A and Substrate Solution B are added and then incubated to produce a blue color indicating the amount of Rubella IgG antibodies present in the specimens. Sulfuric acid solution is added to the microwell plate to stop the reaction producing a color change from blue to yellow. The color intensity, which corresponds to the amount of Rubella IgG antibodies present in the specimens, is measured with a microplate reader at 450/630-700 nm or 450 nm.

Page 2 of 12 Rev. 06, 2017-10-10

MATERIALS PROVIDED

- 1. **Microwell plate**: 12x 8-wells strips coated with purified Rubella antigens
- 2. **Enzyme Conjugate**: 1 vial of 12 mL; Anti-human IgG antibody bound to peroxidase; Preservative: 0.1 % ProClin[™] 300
- 3. **Wash Buffer conc.**: 1 vial of 50 mL; 25x conc., Tris-HCl buffer containing 0.1 % Tween 20; Preservative: 0.1 % ProClin[™] 300
- 4. **Specimen Diluent**: 1 vial of 12 mL; Tris buffer, Preservative: 0.1 % ProClin[™] 300
- Substrate Solution A: 1 vial of 8 mL; Citrate-phosphate buffer containing hydrogen peroxide; Preservative: 0.1 % ProClin[™] 300
- 6. **Substrate Solution B**: 1 vial of 8 mL; Buffer containing tetramethylbenzidine (TMB); Preservative: 0.1 % ProClin[™] 300
- 7. Stop Solution: 1 vial of 8 mL; 0.5 M Sulfuric acid
- 8. **Calibrators**: 4 vials of 1 mL each. Diluted human serum containing the following amounts of Rubella IgG antibodies; Preservative: 0.1 % ProClinTM 300:
 - 1) Calibrator A 0 IU/mL
- 2) Calibrator B 5 IU/mL
- 3) Calibrator C 10 IU/mL
- 4) Calibrator D 200 IU/mL

- 9. Plate sealer
- 10. Package Insert

MATERIALS REQUIRED BUT NOT PROVIDED

- Freshly distilled or deionized water
- Sodium hypochlorite solution for decontamination
- Absorbent paper or paper towel
- Water bath or incubator capable of maintaining 37°C ± 2°C
- Calibrated automatic or manual microwell plate washer capable of aspirating and dispensing 350 µL/well
- Disposable gloves
- Calibrated micropipettes with disposable tips capable of dispensing 5, 50 and 100 μL
- Graduated cylinders for wash buffer dilution
- Vortex mixer for specimen mixing (optional)
- Timer
- Disposable reagent reservoirs
- Calibrated microplate reader capable of reading at 450 nm with a 630-700 nm reference filter, or reading at 450 nm without a reference filter
- Automated processor (optional)

Page 3 of 12 Rev. 06, 2017-10-10

PRECAUTIONS

- For professional in vitro diagnostic use only. Do not use after expiration date.
- Do not mix reagents from other kits with different lot numbers.
- Avoid cross contamination between reagents to ensure valid test results.
- Follow the wash procedure to ensure optimum assay performance.
- Use Plate Sealer to cover microwell plate during incubation to minimize evaporation.
- Use a new pipet tip for each specimen assayed.
- Ensure that the bottom of the plate is clean and dry and that no bubbles are present on the surface of the liquid before reading the plate. Do not allow wells to dry out during the assay procedure.
- Do not touch the bottom of the wells with pipette tips. Do not touch the bottom of the microwell plate with fingertips.
- Do not allow sodium hypochlorite fumes from chlorine bleach or other sources to contact the microwell plate during the assay as the color reaction may be inhibited.
- All equipment should be used with care, calibrated regularly and maintained following the equipment manufacturer's instructions.
- Calibrators, Enzyme Conjugate, Sample Diluent, Substrate Solution A, Substrate Solution B, Wash Buffer:

Above reagents contain 0.1 % ProClin[™] 300 as a preservative, which is classified as below:

H317: May cause an allergic skin reaction.

P272: Contaminated work clothing should not be allowed out of the

workplace.

P280: Wear protective gloves/protective clothing/eye protection/face protection.

P302+P352: If on skin: wash with plenty of soap and water.

P333+P313: If skin irritation or rash occurs: Get medical advice/attention.
P362+P364: Take off contaminated clothing and wash it before reuse.
P501: Dispose of contents and container in accordance to local,

regional, national and international regulations.



HEALTH AND SAFETY INFORMATION

- Some components of this kit contain human blood derivatives, which were found to be non-reactive for the HIV-1/HIV-2/HIV-O, Syphilis and HCV antibodies, as well as HBsAg. But no known test method can offer complete assurance that products derived from human blood will not transmit infectious agents. Therefore, all blood derivatives should be considered potentially infectious. It is recommended that these reagents and human specimens be handled using established good laboratory working practices.
- Wear disposable gloves and other protective clothing such as laboratory coats and eye
 protection while handling kit reagents and specimens. Wash hands thoroughly when
 finished.
- ProClin[™] 300 is included as a preservative in the Conjugate, Concentrated Wash Buffer, Specimen Diluent, Substrate Solutions and Calibrators. Avoid any contact with skin or eyes.
- Do not eat, drink or smoke in the area where the specimens or kits are handled. Do not pipette by mouth.
- Avoid any contact of the Substrate Solution A, Substrate Solution B, and Stop Solution with skin or mucosa. The Stop Solution contains 0.5 M sulfuric acid which is a strong acid. If spills occur, wipe immediately with large amounts of water. If the acid contacts the skin or eyes, flush with large amounts of water and seek medical attention.
- Non-disposable apparatus should be sterilized after use. The preferred method is to autoclave for one hour at 121°C. Disposables should be autoclaved or incinerated. Do not autoclave materials containing sodium hypochlorite.
- Handle and dispose all specimens and materials used to perform the test as if they
 contained infectious agents. Observe established precautions against microbiological

- hazards throughout all the procedures and follow the standard procedures for proper disposal of specimens.
- Observe Good Laboratory Practices when handling chemicals and potentially infectious material. Discard all contaminated material, specimens and reagents of human origin after proper decontamination and by following local, state and federal regulations.
- Neutralized acids and other liquids should be decontaminated by adding sufficient volume of sodium hypochlorite to obtain a final concentration of at least 1.0%. A 30 minute exposure to a 1.0% sodium hypochlorite may be necessary to ensure effective decontamination.

STORAGE AND STABILITY OF THE KIT

- Unopened test kits should be stored at 2-8°C upon receipt. All reagents are stable through the expiration date printed on the box if stored between 2-8°C. Once opened, all reagents are stable for up to 3 months after the first opening date if stored between 2-8°C. Return reagents to 2-8°C immediately after use.
- Allow the sealed pouch to reach room temperature before opening the pouch and removing the required number of strips to prevent condensation of the microwell plate. The remaining unused strips should be stored in the original resealable pouch at 2-8°C and can be used within 3 month of the opening date. Return the remaining unused strips and supplied desiccant to the original resealable pouch, firmly press the seal closure to seal the pouch completely and immediately store at 2-8°C.
- Concentrated Wash Buffer may be stored at room temperature to avoid crystallization. If crystals are present, warm up the solution at 37°C. Working Wash Buffer is stable for 2 weeks at room temperature.
- Do not expose reagents especially the Substrate Solutions to strong light or hypochlorite fumes during storage or incubation steps.
- Do not store Stop Solution in a shallow dish or return it the original bottle after use.

SPECIMEN COLLECTION AND PREPARATION

- The Dialab Rubella IgG ELISA Test Kit can be performed using only human serum or plasma collected from venipuncture whole blood.
- EDTA, sodium heparin, and ACD collection tubes may be used to collect venipuncture whole blood and plasma specimens. The preservative sodium azide inactivates horseradish peroxide and may lead to erroneous results.
- Separate serum or plasma from blood as soon as possible to avoid hemolysis. Grossly hemolytic, lipidic or turbid samples should not be used. Specimen with extensive particulate should be clarified by centrifugation prior to use. Do not use specimens with fibrin particles or contaminated with microbial growth.
- Do not leave specimens at room temperature for prolonged periods. Serum and plasma specimens may be stored at 2-8°C for up to 7 days prior to assaying. For long term storage, specimens should be kept frozen below -20°C.
- Bring specimens to room temperature prior to testing. Frozen specimens must be completely thawed and mixed well prior to testing. Specimens should not be frozen and thawed repeatedly.
- If specimens are to be shipped, they should be packed in compliance with local regulations covering the transportation of etiologic agents.

REAGENTS PREPARATION

WASH BUFFER:

Prepare Working Wash Buffer by diluting the Concentrated Wash Buffer 1:25. Pour the contents of the bottle in a graduated cylinder and fill it with freshly distilled or deionized water to 1250 mL. It is stable for 2 weeks at 15-30°C.

Note: If crystals are present in the Concentrated Wash Buffer, warm it up at 37°C until all crystals dissolve.

Allow reagents and specimens to reach room temperature (15-30°C) prior to testing. The procedure must be strictly followed. Assay must proceed to completion within time limits. Arrange the calibrators so that well A1 is the Blank well. From well A1, arrange the calibrators in a horizontal or vertical configuration. The procedure below assigns specific wells arranged in a vertical configuration. Configuration may depend upon software.

ASSAY PROCEDURE

- 1. Leave A1 as Blank well.
- 2. Add 100 µL of Calibrator 1 in wells B1 and C1. (Yellow Reagent)
 - Add 100 µL of Calibrator 2 in wells D1 and E1. (Blue Reagent)
 - Add 100 µL of Calibrator 3 in wells F1 and G1. (Blue Reagent)
 - Add 100 µL of Calibrator 4 in wells H1 and A2. (Blue Reagent)
- 3. Add 100 μL of Specimen Diluent to assigned wells starting at B2. The color of Specimen Diluent is green.
 - Add 5 μ L of specimen to assigned wells starting at B2. Then a color change from green to blue will occur to verify that the specimen has been added.
 - Remove unused strips from the microwell plate, and store in the original resealable pouch at 2-8°C.
- 4. Mix gently by swirling the microwell plate on a flat bench for 30 seconds.
 - Cover the microwell plate with the Plate Sealer and incubate in a water bath or an incubator at 37°C ± 2°C for 30 minutes ± 2 minutes.
- 5. Remove the Plate Sealer.
 - Wash each well 5 times with 350 μ L of Working Wash Buffer per well, then remove the liquid.
 - Turn the microwell plate upside down on absorbent tissue for a few seconds. Ensure that all wells have been completely washed and dried.
 - Note: Improper washing may cause false positive results.
- 6. Add 100 μ L of Conjugate to each well except for the Blank well. The color of Conjugate is red.
- 7. Cover the microplate plate with the Plate Sealer and incubate in a water bath or an incubator at 37°C ± 2°C for 30 minutes ± 2 minutes.
- 8. Repeat Step 5.
- 9. Add 50 µL of Substrate Solution A to each well. (Clear Reagent)
 - Add 50 µL of Substrate Solution B to each well. (Clear Reagent)
 - Then a blue color should develop in wells containing Positive specimens.
- 10. Mix gently then cover microwell plate with Plate Sealer and incubate in a water bath or incubator at 37°C ± 2°C for 10 minutes ± 1 minute.
- 11. Remove the Plate Sealer.
 - Add 50 µL of Stop Solution to each well. (Clear Reagent)
 - Then a yellow color should develop in wells containing Positive specimens.
- 12. Read at 450/630-700 nm in 30 minutes.
 - Note: Microwell plate can also be read at 450 nm, but it is strongly recommended to read it at 450/630-700 nm for better results.

ASSAY SCHEME

1. Prepare the Working Wash Buffer by diluting the Wash Buffer concentrate 1:25.

2. Follow this scheme:

| REAGENTS | A1 BLANK | CALIBRATORS | SAMPLE | | |
|---|----------|-------------|--------|--|--|
| Calibrators | - | 100 μL | - | | |
| Sample Diluent | - | - | 100 µL | | |
| Sample | - | - | 5 μL | | |
| Cover strips with adhesive film. | | | | | |
| Incubate 30 min. at +37°C. | | | | | |
| Peel out the adhesive film and aspirate the reaction solution from all wells. | | | | | |
| Wash 5 times with 350 uL of diluted Wash Buffer, carefully aspirating off the remaining liquid. | | | | | |

| Enzyme Conjugate | - | 100 μL | 100 μL | | |
|---|---------------------|----------------------|--------|--|--|
| Cover strips with adhesive film. | | | | | |
| Incubate 3 | 30 min. at +37°C. | | | | |
| Peel out the adhesive film and as | oirate the reaction | solution from all we | ells. | | |
| Wash 5 times with 350 µL of diluted Wash Buffer, carefully aspirating off the remaining liquid. | | | | | |
| Substrate Solution A 50 µL 50 µL 50 µL | | | | | |
| Substrate Solution B | 50 μL | 50 μL | 50 μL | | |
| Cover strips with a new adhesive film. | | | | | |
| Incubate 10 min. at +37°C., protected from light. | | | | | |
| Stop solution | 50 μL | 50 μL | 50 μL | | |
| Read the absorbance of each well against A1 blanking-well at 450 nm and 630-700 nm in 30 min. | | | | | |

AUTOMATED PROCESSING

Automatic ELISA microplate processors may be used to perform the assay after validating the results to ensure they are equivalent to those obtained using the manual method for the same specimens. Incubation times may vary depending on the processors used but do not program less incubation times than the procedure listed above. When automatic ELISA microplate processors are used, periodic validation is recommended to ensure proper results.

VALIDATION REQUIREMENTS AND QUALITY CONTROL

1. Calculate the Mean Absorbance of Calibrators 1-4 by referring to the table below.

Example of Calibrator 3 Calculation

| Item | Absorbance | |
|----------------------------------|-----------------------|--|
| Calibrator 3: Well F1 | 1.012 | |
| Calibrator 3: Well G1 | 1.102 | |
| Total Absorbance of Calibrator 3 | 1.012 + 1.102 = 2.114 | |
| Mean Absorbance of Calibrator 3 | 2.114/2 = 1.057 | |

2. Check the validation requirements below to determine if the test results are valid.

| Item | Validation Requirements | | |
|--------------|---|--|--|
| Blank Well | Blank Absorbance should be < 0.05 if read at 450/630-700 nm | | |
| | Note: It should be < 0.100 if read at 450 nm | | |
| Calibrator 1 | Mean Absorbance < 0.100 after subtraction of Blank Absorbance | | |
| Calibrator 2 | Mean Absorbance > 0.200 and < 0.700 after subtraction of Blank Absorbance | | |
| Calibrator 3 | Mean Absorbance > Calibrator 2 and < Calibrator 4 after subtraction of Blank Absorbance | | |
| Calibrator 4 | Mean Absorbance > 1.500 after subtraction of Blank Absorbance | | |

NOTE: The test results are considered invalid if the above validation requirements are not met. Repeat the test or contact your local distributor.

INTERPRETATION OF RESULTS

Qualitative

Calculate the Index Value to obtain qualitative specimen results.

1. If the test is valid, obtain Cut-Off Value by subtracting the Blank Absorbance from the Mean Absorbance of Calibrator 3. See an example of Cut-Off Value calculation below.

| Item | Absorbance | |
|---|-----------------------|--|
| Blank Absorbance: Well A1 | 0.014 | |
| Cut-Off Value: Mean Absorbance of Calibrator 3 – Blank Absorbance | 1.057 – 0.014 = 1.043 | |

2. Calculate the Index Value by dividing the Specimen Absorbance by the Cut-Off Value, then read the results by referring to the Interpretation of Results table below

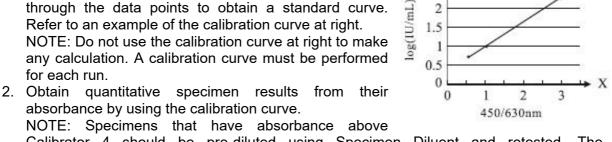
| then read the results by releming to the in | terpretation of results table below. | |
|---|--------------------------------------|--|
| ltem | Absorbance | |

| Specimen: Well B2 | 1,779 | | |
|-------------------------------------|---------------------|--|--|
| Cut-Off Value | 1,043 | | |
| Index Value: Specimen/Cut-Off Value | 1.779/1.043 = 1.710 | | |

Quantitative

Draw the calibration curve and obtain quantitative specimen results.

- 1. Subtract the Blank Absorbance from the Mean Absorbance of each Calibrator, then plot
 - them on the X-axis against their concentration in IU/mL on the Y-axis on a semi-logarithmic graph paper and draw the calibration curve. Draw the best fitted line through the data points to obtain a standard curve. Refer to an example of the calibration curve at right.



2

Calibrator 4 should be pre-diluted using Specimen Diluent and retested. The concentration must be multiplied by the dilution factor. Automated reading and calculation may be performed using linear regression function on suitable computer programs.

Interpretation of Results – Qualitative and Quantitative

| Results | Qualitative | Quantitative | |
|------------|-----------------|-----------------|--|
| Results | Index Value | Concentration | |
| Negative | < 0.5 | < 5.0 U/mL | |
| Positive | > 1.1 | ≥ 10.0 U/mL | |
| Equivocal* | ≥ 0.5 and ≤ 1.1 | 5.0 – 10.0 U/mL | |

*NOTE: For Equivocal results, the specimen should be retested. Specimens that are repeatedly Equivocal after retest should be confirmed using an alternate method. If the results remain Equivocal, collect a new specimen in two weeks. If the new specimen is Positive, the specimen is presumed to be Positive.

LIMITATIONS

- 1. The Dialab Rubella IgG ELISA Test Kit is used for the detection of IgG antibodies to Rubella in human serum or plasma. Diagnosis of an infectious disease should not be established based on a single test result. Further testing, including confirmatory testing, should be performed before a specimen is considered positive. A negative test result does not exclude the possibility of exposure. Specimens containing precipitate may give inconsistent test results.
- 2. As with all diagnostic tests, all results must be interpreted together with other clinical information the physician.
- 3. As with other sensitive immunoassays, there is the possibility that the positive result cannot be repeated due to inadequate washing from the initial test. The results may be affected due to procedural or instrument error.

PERFORMANCE CHARACTERISTICS

The calibrators are referenced to the World Health Organization International Standard for Anti-Rubella Serum (3rd International Standard Preparation) at each concentration level.

Sensitivity and Specificity

The Dialab Rubella IqG ELISA Test Kit has correctly identified specimens of a mixed titer performance panel and has been compared to a leading commercial Dialab Rubella IgG ELISA test using clinical specimens. The results show that the clinical sensitivity of the Dialab Rubella IgG ELISA Test Kit is 96.4%, and the clinical specificity is >99.9%.

Dialab Rubella IgG ELISA vs. Other ELISA

| Method | | Other ELISA | | Total Doculto | |
|----------------------|----------|-------------|----------|---------------|--|
| | Results | Positive | Negative | Total Results | |
| Rubella IgG ELISA | Positive | 54 | 0 | 54 | |
| ELISA | Negative | 2 | 37 | 39 | |
| Total Results | | 56 | 37 | 93 | |

Clinical Sensitivity: 96.4% (87.7-99.6%)* Clinical Specificity: >99.9% (90.5-100.0%)* Overall Agreement: 97.9% (92.4-99.7%)*

REPRODUCIBILITY

Intra-Assay: Within-run precision has been determined by using 10 replicates of three specimens: a low positive, a medium positive and a strong positive.

Inter-Assay: Between-run precision has been determined by 3 independent assays on the same three specimens: a low positive, a medium positive and a strong positive. Three different lots of the Dialab Rubella IgG ELISA Test Kit have been tested using these specimens over a 5-day period.

| | Intra-Assay | | | Inter-Assay | | |
|----------|---------------------------------|-----------------------|------------------------------|---------------------------------|-----------------------|------------------------------|
| Specimen | Mean Absorbance / Cut-Off | Standard Deviation | Coefficient of Variation (%) | Mean Absorbance / Cut-Off | Standard Deviation | Coefficient of Variation (%) |
| 1 | 0,508 | 0,018 | 3,528 | 0,508 | 0,029 | 5,786 |
| 2 | 1,206 | 0,065 | 5,390 | 1,229 | 0,057 | 4,638 |
| 3 | 1,884 | 0,111 | 5,892 | 1,846 | 0,111 | 6,013 |

INTERFERENCES

Interferences are not observed up to a concentration of 1mg/mL Acetaminophen, 0.2 mg/mL Gentistic Acid, 0.1 mg/mL Ascorbic Acid, 0.1 mg/mL Acetosalisilyc Acid, 0.1 mg/mL Caffeine, 0.6 mg/mL Oxalic Acid, 2 mg/mL Bilirubin, 2 mg/mL Hemoglobin, 10% H₂O₂, 1 % Methanol and 1% Ethanol. Rheumatoid factors do not interfere with the test.

Cross-reactivity is not observed in Syphilis, HBsAg, HIV, HCV, HSV 1 IgG, Toxoplasma IgG and CMV IgG positive specimens.

REFERENCES

1. Herrmann, KL. Rubella Virus. In: Manual of Clinical Microbiology. American Society for

Page 9 of 12 Rev. 06, 2017-10-10

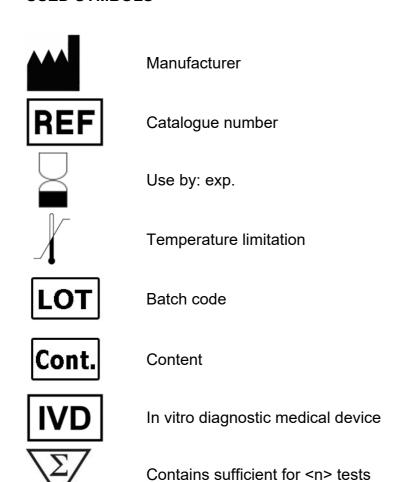
^{*95%} Confidence Interval

Rubella IgG

- Microbiology 4th Edition (1985) 779-784.
- 2. Turgeon, ML. Rubella Infection. In: Immunology and Serology in Laboratory Medicine. 2nd Edition (1996). 275-286.
- 3. Chernesky, MA, Mahony JB. Rubella Virus. In: Manual of Clinical Microbiology. 6th Edition (1995) 968-973.
- 4. Voller, A, Bidwell, DE, A simple Method for Detecting Antibodies to Rubella. Brit. J. Exp. Pathol. (1975) 56:338-339.
- 5. Rawls WE, Chernesky MA. Rubella Virus. Manual Clinical Immunology (1976) 452-455.
- 6. Millian, SJ, Wegman D. Rubella Serology: Applications, Limitations, and Interpretations. Amer. J. Pub. Health (1972) 170-176.

Page 10 of 12 Rev. 06, 2017-10-10

USED SYMBOLS



ELISA Enzyme Linked Immunosorbent Assay



DIALAB Produktion und Vertrieb von chemisch – technischen Produkten und Laborinstrumenten Gesellschaft m.b.H.

IZ-NÖE Sued, Hondastrasse, Objekt M55, A – 2351 Wiener Neudorf, Austria Phone: ++43 (0) 2236 660910-0, Fax: ++43 (0) 2236 660910-30, e-mail: office@dialab.at